

2-Ethyl-2-phosphabicyclo[2.2.2]oct-7-ene Derivatives: Synthesis and Use in Fragmentation-Related Phosphorylations

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ABSTRACT: A 2-phosphabicyclo[2.2.2]oct-7-ene oxide (**2**) and a 2-phosphabicyclo[2.2.2]octa-5,7-diene oxide (**3**) with ethyl substituent on the phosphorus atom was synthesized and their fragmentation properties were studied. The phosphabicyclooctadiene oxide (**3**) could be utilized in both the UV light-mediated phosphorylation of simple alcohols and in the thermoinduced phosphorylation of hydroquinone giving an easy access to P-ethylphosphinates (e.g., **4** and **6**). The phosphabicyclooctene oxide (**2**) was, however, not useful in photoinduced phosphorylations; under such conditions the precursor (**2**) underwent dechlorination to afford **5**. © 2005 Wiley Periodicals, Inc. *Heteroatom Chem* 16:196–199, 2005; Published online in Wiley InterScience (www.interscience.wiley.com). DOI 10.1002/hc.20093

INTRODUCTION

The bridged P-heterocycles, such as the 2-phosphabicyclo[2.2.2]octadiene 2-oxides/sulfides can be regarded as precursors of methylenephosphine oxides/sulfides $YP(X)CH_2$, where $X = O, S$, $Y = \text{phenyl, alkoxy}$. The latter are novel phosphorylating agents toward a variety of nucleophiles, such as alcohols, phenols, and amines [1,2]. The reaction can be achieved either thermally [3], or under photochemical conditions [4–6]. In the latter instance, a novel addition-elimination mechanism was substantiated to be a competitive pathway [5–7] in addition to the traditional elimination–addition route involving methylenephosphine oxide as the intermediate [4]. The fragmentation-related phosphorylations are of interest, as they require mild reaction conditions, especially those accomplished under photochemical conditions, (26°C and an irradiation of only several hours) and take place efficiently and in a selective way [5,6,8]. During our efforts to find suitable precursors and to evaluate the mechanism of the fragmentations, a series of novel phosphabicyclooctene derivatives were described [9–11]. To extend the sphere of the precursors available, bridged P-heterocycles with ethyl substituent on the phosphorus atom are introduced and their

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fragmentation properties are described in the present paper.

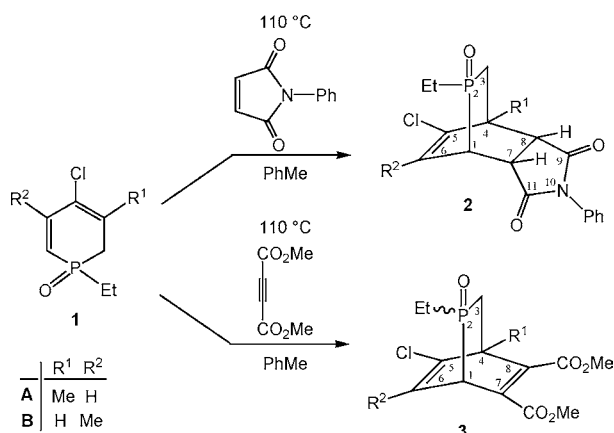
RESULTS AND DISCUSSION

Double-bond isomers of the 1-ethyl-1,2-dihydrophosphinine oxide (**1A** and **1B**) [12] were reacted with *N*-phenylmaleimide in boiling toluene to give phosphabicyclo[2.2.2]octene oxide **2** as the mixture of regioisomers (**A** and **B**) (Scheme 1). Similarly, Diels–Alder reaction of the mixture of **1A** and **1B** with dimethyl acetylenedicarboxylate afforded the regioisomers (**A** and **B**) of phosphabicyclo[2.2.2.]octadiene oxide **3**. In this instance, regioisomers **A** and **B** consisted of configurational isomers (Scheme 1).

The crude products **2** and **3** were purified by column chromatography to provide the cycloadducts **2** and **3** in 74% and 54%, yield, respectively. Isomers of the products **2** and **3** were characterized by ^{31}P , ^{13}C , and ^1H NMR, as well as mass spectrometry.

Thermal examinations (TG, DTG, and DSC) suggested that phosphabicyclooctene oxide **2** underwent the elimination of the bridging P-moiety in the range of 270–430°C, with a minimum in the DSC at 380°C. This revealed a considerably larger thermostability, as compared to that of phosphabicyclooctadiene oxide **3** eliminating the bridging unit in the range of 190–320°C with a minimum in the DSC at 257°C. Due to their more strained ring, the phosphabicyclooctadiene oxides of type **3** are indeed thermally less stable [13,14].

In the next stage of our work, we tested the new precursors **2** and **3** in fragmentation-related phosphorylations. Irradiation of the acetonitrile solution of phosphabicyclooctadiene oxide **3** at 254 nm in the presence of methanol or ethanol at 26°C yielded the



SCHEME 1

corresponding *P*-ethyl phosphinates **4a** and **4b**, respectively in ca. 71% yield and in a purity of ca. 95% after flash column chromatography (Scheme 2).

Photolysis of phosphabicyclooctene oxide **2** under similar conditions using methanol did not give, however, the expected phosphinate **4a**. Only traces of the desired product **4a** could be detected. Instead, precursor **2** underwent dechlorination to furnish phosphabicyclooctene oxide **5** (Scheme 3).

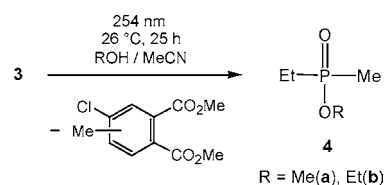
Like cycloadduct **2**, compound **5** formed also did not eliminate the bridging P-unit. The unreactivity of phosphabicyclooctene oxides **2** and **5** is surprising, as on the basis of the UV absorption they should be suitable precursors. The UV spectrum of cycloadduct **2** showed an intensive absorption in the range of ~220–270 nm with a maximum at 252 nm that is similar to the spectrum of the *P*-phenyl analogue revealing a maximum at 254 nm.

Phosphabicyclooctadiene oxide **3** was also useful in the thermo-induced phosphorylation of hydroquinone. Heating the mixture of **3** and the hydroxy-compound at 240°C for 10 min led to phosphinate **6** in 64% yield and in a purity of 94% after flash column chromatography (Scheme 4). Carrying out the fragmentation-related phosphorylation under microwave conditions, the reaction was more efficient (72% yield), and the product (**6**) was cleaner (98%). Phosphinates **4a**, **4b**, and **6** were characterized by ^{31}P NMR and mass spectrometrical data. Mass spectral data were obtained by GC-MS.

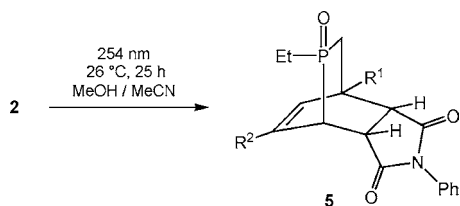
To summarize the results, two new phosphabicyclo[2.2.2]octene derivatives with ethyl substituent at the phosphorus atom were introduced and tested in thermoinduced and UV light-mediated fragmentation-related phosphorylations. Hence, our methodology has now been extended to *P*-ethyl model compounds.

EXPERIMENTAL

The ^{31}P , ^{13}C , and ^1H NMR spectra were recorded on a Bruker DRX-500 spectrometer operating at 202.4, 125.7, and 500 MHz, respectively. Chemical shifts are downfield relative to 85% H_3PO_4 or TMS. The couplings are given in Hz. GC-MS was



SCHEME 2



SCHEME 3

performed on a Fisons GC 8000/MD 800 apparatus connecting a PE SCIEX API 2000 type triple quadrupol mass spectrometer. FAB mass spectrometry was performed on a ZAB-2SEQ instrument. The ethyl-dihydrophosphinine oxides (**1A** and **1B**) were prepared as described earlier [12].

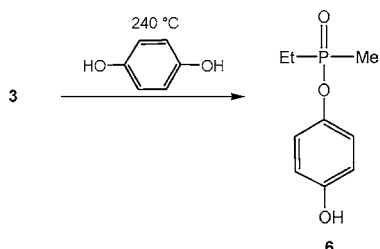
General Procedure for the Synthesis of Cycloadducts **2** and **3**

The mixture of 1.0 g (5.25 mmol) of ethyl-dihydrophosphinine oxide **1** consisting of 75% of the **A** isomer and 25% of the **B** isomer and 5.78 mmol of the dienophile (1.0 g of *N*-phenylmaleimide or 0.72 mL of dimethylacetylene dicarboxylate) in 25 mL of toluene was stirred at the boiling point for 5 days. Solvent was evaporated, and the residue so obtained was purified by column chromatography (silica gel, 3% methanol in chloroform) to furnish cycloadduct **2** and **3**, respectively, as the mixture of isomers. The data are listed below.

1- and 11-Methyl-10-chloro-4-phenyl-8-ethyl-4-aza-8-phosphatricyclo[5.2.2.0^{2,6}]undec-10-ene-3,5-dione 8-oxide (**2A** and **2B**) [15]

Yield: 1.4 g (74%) of **2** as a 6:4 mixture of isomers **A** and **B**; mp 257–259°C (acetone); FAB-MS, 364 (*M* + *H*); (*M* + *H*)_{found}⁺ = 364.0869, C₁₈H₂₀ClNO₃P requires 364.0806 for the ³⁵Cl isotopomer.

2A: ³¹P NMR (CDCl₃) δ 51.8; ¹³C NMR (CDCl₃) δ 5.1 (²*J* = 5.3, CH₂CH₃), 21.4 (¹*J* = 70.9, CH₂CH₃), 23.3 (³*J* = 10.3, C(4)–CH₃), 34.8 (¹*J* = 70.3, C(3)), 58.7 (¹*J* = 36.6, C(1)), 38.9 (C(7)), 43.8 (²*J* = 6.1,



SCHEME 4

C(4)), 49.2 (²*J* = 10.1, C(8)), 121.8 (²*J* = 5.0, C(6), 126.3 (C(3'))*, 128.6 (C(4')), 128.9 (C(2'))*, 131.4 (C(1')), 139.5 (³*J* = 10.8 C(5)), 173.9 (C(9)), 175.9 (³*J* = 14.1, C(11)), * may be reversed; ¹H NMR (CDCl₃) δ 2.38 (m, CH₂CH₃), 2.85 (s, C(4)–CH₃), 7.27 (dd, ³*J*_{PH} = ³*J*_{HH} 7.7, C(6)–H).

2B: ³¹P NMR (CDCl₃) δ 58.7; ¹³C NMR (CDCl₃) δ 5.1 (²*J* = 5.3, CH₂CH₃), 18.3 (C(6)–CH₃), 21.3 (¹*J* = 69.8, CH₂CH₃), 26.4 (¹*J* = 71.9, C(3)), 39.7 (C(7)), 41.6 (¹*J* = 58.2, C(1)), 42.5 (²*J* = 6.9, C(4)), 45.0 (²*J* = 11.9, C(8)), 126.2 (C(3'))*, 139.5 (³*J* = 11.0, C(5)), 128.7 (C(4')), 129.0 (C(2'))*, 129.8 (²*J* = 5.0, C(6)), 131.4 (C(1')), 175.1 (C(9)), 176.1 (³*J* = 13.8, C(11)), * may be reversed; ¹H NMR (CDCl₃) δ 2.38 (m, CH₂CH₃), 2.99 (s, C(6)–CH₃).

4- and 7-Methyl-8-chloro-2-ethyl-2-oxo-2-phosphabicyclo[2.2.2]octa-5,7-diene-5,6-dicarboxylic Acid Dimethyl Ester (**3**) [15]

Yield: 0.94 g (54%) oily product after repeated chromatography (silica gel, 3% methanol in chloroform) as a 32%, 29%, 25%, and 14% mixture of four isomers; FAB-MS, 333 (*M* + *H*); (*M* + *H*)_{found}⁺ = 333.0658, C₁₄H₁₉ClO₅P requires 333.0601 for the ³⁵Cl isotopomer.

3A-1: ³¹P NMR (CDCl₃) δ 58.9 (32%); ¹³C NMR (CDCl₃) δ 5.6 (CH₂CH₃)^a, 19.7 (*J* = 9.4, C(4)–CH₃)^b, 30.8 (*J* = 89.8, CH₂)^c, 34.0 (*J* = 109.9, C(3))^d, 41.5 (*J* = 46.8, C(1))^e, 47.0 (*J* = 8.3, C(4))^f, 52.4^g, 52.3^g (MeO), 124.2 (*J* = 11.5, C(6))^h, 136.4 (*J* = 5.7, C(7))ⁱ, 138.3 (*J* = 19.2, C(8))^j, 162.2 (*J* = 4.6, C(10))^k, 164.3 (C(9))^l.

3A-2: ³¹P NMR (CDCl₃) δ 59.6 (29%); ¹³C NMR (CDCl₃) δ 5.5 (CH₂CH₃)^a, 19.6 (*J* = 8.7, C(4)–CH₃)^b, 30.3 (*J* = 89.6, CH₂)^c, 33.5 (*J* = 103.5, C(3))^d, 41.2 (*J* = 46.9, C(1))^e, 46.5 (*J* = 8.2, C(4))^f, 51.9^g, 52.0^g (MeO), 123.9 (*J* = 6.8, C(6))^h, 134.6 (*J* = 6.3, C(7))ⁱ, 139.4 (*J* = 18.6, C(8))^j, 163.9 (C(9))^l, 165.6 (*J* = 3.2, C(10))^k, ^{a–l} tentative assignment.

3B-1: ³¹P NMR (CDCl₃) δ 55.3 (25%); ¹³C NMR (CDCl₃) δ 5.7 (CH₂CH₃), 18.1 (C(6)–CH₃), 48.2 (*J* = 46.1, C(1)), 45.0 (*J* = 8.9, C(4)), 52.2, 52.4 (MeO), 130.9 (*J* = 10.9, C(6)), 132.0 (C(7)), 139.7 (*J* = 16.3, C(8)).

3B-2: ³¹P NMR (CDCl₃) δ 55.5 (14%).

General Procedure for the Photoinduced Fragmentation-Related Phosphorylations

The solution of 0.10 g (0.301 mmol) of precursor **3** as a 32, 29, 25, and 14% mixture of isomers (obtained as shown above) in 45 mL of dry acetonitrile was irradiated in the presence of 4.0 mL of the corresponding alcohol (methanol or ethanol) in a photochemical

quartz reactor with a 125 W mercury lamp for 2 h. Volatile components were removed, and the residue so obtained purified by flash column chromatography (silica gel, 3% methanol in chloroform) to give the corresponding phosphinic esters (**4a** and **4b**) as oils in a purity of ca. 95%.

4a was obtained in the presence of methanol; yield: 74%; ^{31}P NMR (CDCl_3) δ 59.0; ES-MS, 123 ($\text{M} + \text{H}$); HR-FAB ($\text{M} + \text{H}$) $^+$ _{found} = 123.0555, $\text{C}_4\text{H}_{11}\text{O}_2\text{P}$ requires 123.0575; GC-MS, m/z (rel. int.) 122 (M^+ , 28), 107 (22), 94 (100), 91 (5), 79 (87), 59 (61).

4b was obtained in the presence of ethanol; yield: 69%; ^{31}P NMR (CDCl_3) δ 57.4; ES-MS, 137 ($\text{M} + \text{H}$); HR-EI M^+ _{found} = 136.0632, $\text{C}_5\text{H}_{13}\text{O}_2\text{P}$ requires 136.0653; GC-MS, m/z (rel. int.) 136 (M^+ , 70), 135 (100), 107 (17), 92 (14), 77 (30).

Photolysis of phosphabicyclooctene regioisomers **2A** and **2B** under similar conditions in the presence of methanol led to the dechlorinated derivative of precursor **5**. After an irradiation time of 75 min, the conversion was found to be 53%. ^{31}P NMR (CDCl_3) δ 55.2; FAB-MS, 330 ($\text{M} + \text{H}$).

Thermoinduced Fragmentation-Related Phosphorylation of Hydroquinone Using Phosphabicyclooctadiene 3

0.20 g (0.60 mmol) of cycloadduct **3** as the mixture of four isomers and 0.20 g (1.8 mmol) of hydroquinone was heated at 240°C in a vial for 10 min. Flash column chromatography (silica gel, 3% methanol in chloroform) afforded 7.7 mg (64%) phosphinate **6** in a purity of ca. 94%. ^{31}P NMR (CDCl_3) δ 59.3; FAB-MS, 201 ($\text{M} + \text{H}$); HR-FAB ($\text{M} + \text{H}$) $^+$ _{found} = 201.0647, $\text{C}_9\text{H}_{13}\text{O}_3\text{P}$ requires 201.0681; GC-MS, m/z (rel. int.) 200 (M^+ , 100), 185 (1), 172 (12), 171 (10), 157 (37), 110 (80), 107 (57), 91 (51).

Microwave-Assisted Fragmentation-Related Phosphorylation of Hydroquinone Using Phosphabicyclooctadiene 3

A mixture of cycloadduct **3** (0.10 g, 0.30 mmol) and 0.10 g (0.9 mmol) of hydroquinone in a glass vial was placed into a focused 300 W CEM Discover microwave reactor, and the sample was heated at 240°C for 10 min with a power of 35 W in the stationer stage. Flash column chromatography (silica gel, 3% methanol in chloroform) afforded 8.6 mg (72%) phosphinate **6** in a purity of ca. 98%.

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- [15] For naming compounds **2** and **3**, the IUPAC nomenclature was used (see below). At the same time, the traditional numbering was applied during the NMR assignments.

